

Soil Salinity Near Rivers, Lakes, and Oceans Lab Experiment

Posted on September 19, 2018

Pre-Lab Experiment

I am going to carry out an experiment to compare the salinity of soils collected near three different types of water bodies: a permanent river, a freshwater lake, and the ocean. The purpose is to determine whether the location near a water body is associated with measurable differences in soil salinity and to consider how those differences may affect the crops and natural vegetation that can grow along the banks or shoreline. The original plan correctly recognizes that salts in water can influence nearby soils, but location alone will not determine the result. Bedrock, evaporation, rainfall, drainage, irrigation, tidal flooding, distance from the water, elevation, soil texture, and human activity can all affect the amount of soluble salt present.

The experiment will measure salinity through electrical conductivity rather than relying on appearance or taste. Dissolved ions conduct electricity, so an extract prepared from a saline soil generally has higher conductivity than an extract from a soil containing fewer soluble salts. Electrical conductivity does not identify every individual ion, but it provides a practical measurement of total salt concentration. The study will also record soil pH because salinity and acidity are different properties and may influence plants in different ways (Corwin & Yemoto, 2017).

Research Question

How does the electrical conductivity of soil collected near a river, freshwater lake, and ocean differ when samples are collected and analyzed through the same procedure? A second question is whether the measured salinity values are consistent with the types of plants likely to tolerate each location. The experiment is comparative and exploratory. It cannot prove that the water body alone caused the measured soil properties because the sites are not identical in geology, vegetation, drainage, or human disturbance.

Hypothesis

My hypothesis is that soil collected close to the ocean will have the highest electrical conductivity because seawater contains a high concentration of dissolved salts and coastal soils may receive salt through tides, storm surge, sea spray, shallow saline groundwater, or evaporation. I expect the river soil to have the lowest conductivity if the river is freshwater and the bank is well drained. Lake soil

may fall between the two, but its salinity will depend strongly on whether the lake has an outlet, the mineral composition of its watershed, and the local rate of evaporation. A closed lake in an arid region can be more saline than some coastal soils, so the expected order is not universal.

Variables

The independent variable is the sampling location or water-body type. The dependent variables are soil electrical conductivity and pH. Controlled variables should include the mass of soil, volume and quality of extraction water, mixing time, settling time, temperature during measurement, instrument calibration, sampling depth, approximate distance from the water, and container type. Additional observations should include moisture, texture, vegetation, recent rainfall, tidal condition, visible salt crust, and evidence of fertilizer or pollution.

Introduction

Salinity is the concentration of soluble salts in water or soil. Common ions include sodium, calcium, magnesium, potassium, chloride, sulfate, bicarbonate, and carbonate. Seawater has an average salinity of about 35 parts per thousand, although the value varies by location and depth. Freshwater generally contains far less dissolved salt, but it should not be assigned one universal value below 0.1 without identifying the measurement unit. Soil salinity is commonly evaluated by measuring the electrical conductivity of an extract, especially the saturated paste extract, and reporting the result in decisiemens per meter (Corwin & Yemoto, 2017; United States Salinity Laboratory Staff, 1954).

Salinity affects plants by lowering the osmotic potential of soil water. Even when the soil appears moist, a plant may have difficulty extracting water because dissolved salts hold water more strongly. This condition is sometimes described as physiological drought. Particular ions can also become toxic, and high sodium can damage soil structure by dispersing clay, reducing infiltration, and creating crusting. Crops differ greatly in tolerance. Barley and cotton tolerate more salt than beans or many fruit crops, while coastal plants may possess adaptations that allow them to survive saline conditions (United States Salinity Laboratory Staff, 1954).

Soil Collection

I will collect soil from the bank of a permanent freshwater river, the shore of a freshwater lake, and a coastal site near the ocean. Samples should not be taken directly from submerged sediment unless the research question specifically concerns the bed of each water body. Comparing riverbed mud, lake sediment, and dry coastal soil would introduce major differences in moisture and sedimentation. For a fairer comparison, I will collect shoreline or bank soil at a consistent distance from the normal waterline and from a consistent depth, such as the upper 0-10 centimeters after removing loose surface litter.

At least three separate samples should be collected from each location rather than one container from each site. Replicates help show whether a result is typical or caused by one unusual patch. The sampling points should be spaced apart, and each container should be labeled with site, replicate number, date, time, distance from water, and depth. Clean tools should be used between samples to prevent salt from one site contaminating another.

The original procedure stores the samples in sealed plastic containers and keeps the laboratory at constant temperature. Sealing is useful during transport, but very wet soil should not remain sealed for long before processing because biological and chemical conditions can change. Samples may be air-dried according to the selected method, gently broken apart, and passed through an appropriate sieve. The preparation procedure must be identical for all samples.

Soil pH

Soil pH measures hydrogen-ion activity and indicates whether soil is acidic, neutral, or alkaline. It does not directly measure salinity. Many crops grow well within a moderately acidic to neutral range, often around pH 6 to 7.5, but different plants and soil organisms have different preferences. The original statement that most organisms survive only between pH 5 and 6 is too narrow. Strong acidity or alkalinity can reduce nutrient availability and alter microbial communities, but healthy soils occur across a wider range.

Rainwater is naturally slightly acidic because carbon dioxide dissolves to form carbonic acid. Air pollution can add sulfur and nitrogen compounds that form stronger acids, but these should not be described simply as “carbon ions and sulfur.” Acid rain may lower pH in soils with limited buffering capacity. Coastal soils can be alkaline, acidic, or neutral depending on minerals, organic matter, drainage, and oxidation of sulfide-containing sediments.

Temperature and Aquatic Organisms

Water and soil temperature are influenced by sunlight, air temperature, season, depth, shade, currents, groundwater, and soil properties. Wind can mix surface water and change heat distribution. Plants shade soil and can reduce daytime temperature, while organic matter and moisture influence heat storage. Soil organisms produce some metabolic heat, but under ordinary field conditions they do not control temperature in the same way as solar radiation and weather.

The earlier claim that catfish raise water temperature by emitting approximately 35 degrees Celsius is incorrect. Fish are ectothermic and their body temperature generally follows the surrounding water. They produce metabolic heat, but they do not heat an ordinary river or lake to a meaningful degree. The presence of fish may indicate suitable temperature and oxygen conditions rather than cause those conditions.

How Water Bodies Can Affect Soil Salinity

Ocean water can increase coastal soil salinity through direct flooding, spray, tidal exchange, and saline groundwater. Evaporation leaves dissolved salts behind, especially in warm and dry environments. Heavy rain and good drainage may leach salts downward and reduce surface salinity. The distance and elevation of the sampling site therefore matter.

Riverbank soils may receive minerals carried by water and sediments. Flooding can deposit both fertile silt and soluble salts. In humid freshwater systems, regular flow and leaching may keep salinity low. In arid regions or downstream of irrigation, river water can carry enough dissolved salt to increase soil salinity.

Lake-shore soils vary widely. A lake with an outlet can remain fresh because water carries dissolved salts away. A closed basin loses water mainly through evaporation, allowing salts to accumulate. Underlying geology also influences the ions entering the lake. Therefore, the proposed experiment should not assume that all lake soils have medium salinity.

Possible Outcomes

The most likely outcome for the selected local sites is that ocean-adjacent soil will show the highest conductivity, lake-shore soil an intermediate value, and riverbank soil the lowest. This pattern would support the hypothesis but would not prove that ocean animals released the salts. Most ocean salts originate from long-term weathering of rocks, river inputs, hydrothermal processes, and cycling within the Earth system. Coastal accumulation occurs through seawater contact and evaporation rather than primarily through creatures releasing salt.

An alternative result is possible. The lake sample may have the highest conductivity if the lake is saline or receives agricultural drainage. A fertilized riverbank may have elevated conductivity because fertilizers contain soluble ions. Ocean soil collected above the reach of spray and tides after heavy rainfall may have a lower reading than expected. Unexpected results should be investigated rather than forced to match the hypothesis.

The conclusion about fertility also requires caution. Low salinity can support many crops, but riverbank soil is not automatically more fertile. Fertility depends on nutrients, organic matter, texture, pH, drainage, contamination, and flooding risk. A soil can have low salinity yet lack nitrogen or phosphorus. A saline soil can support salt-tolerant vegetation while being unsuitable for sensitive crops.

Materials and Methods

Materials

The experiment will require nine clean sample containers, a clean stainless-steel scoop or soil auger, labels, permanent marker, disposable gloves, sealable bags, a balance, distilled or deionized water, graduated cylinders, mixing containers, stirring rods, filter paper or extraction equipment, an electrical-conductivity meter, a pH meter or calibrated pH probe, calibration solutions, thermometer, sieve, and a laboratory notebook. A camera or phone can document the sites if permitted.

Sampling Method

I will use stratified site sampling with replication rather than claiming that a few samples represent every river, lake, and ocean shoreline. Each water-body type forms one sampling category. Within each category, three points will be selected at a similar distance from the water and sampled at the same depth. The three replicates will be analyzed separately so that variation can be calculated.

Random selection within an accessible shoreline segment can reduce personal choice, but safety and site permission take priority. Sampling points should avoid visible trash, animal waste, road salt piles, fertilizer spills, and drainage outlets unless those influences are part of the question. Notes should record any unavoidable disturbance.

Electrical Conductivity Procedure

The preferred laboratory reference method is electrical conductivity of the saturated paste extract. Water is added gradually to a measured soil portion until a saturated paste with a characteristic consistency forms. The paste is allowed to equilibrate, and the liquid extract is obtained by suction or filtration. The conductivity meter is calibrated and used to measure the extract at a controlled or temperature-corrected condition. This value is commonly called EC_e (U.S. Department of Agriculture, Agricultural Research Service, 2017; United States Salinity Laboratory Staff, 1954).

If saturated-paste equipment is unavailable, a fixed soil-to-water extraction ratio can be used, such as 1:2 or 1:5 by mass or volume, provided the same ratio and method are used for every sample. Results from different extraction ratios are not directly interchangeable, so the method must be reported with the value. For each sample, I will weigh the same amount of prepared soil, add the same volume of deionized water, mix for a fixed time, allow settling, filter or decant consistently, and measure conductivity (Corwin & Yemoto, 2017).

Each extract should be measured at least twice. The probe should be rinsed with deionized water between samples and gently dried according to manufacturer instructions. Calibration should be

checked before and after the session. Temperature will be recorded because conductivity changes with temperature (U.S. Department of Agriculture, Agricultural Research Service, 2017).

pH Procedure

Soil pH will be measured using a consistent soil-to-water suspension and a calibrated pH meter. The same prepared samples can be used if the method is appropriate, but pH and conductivity readings should be recorded separately. The probe must be rinsed between samples. Calibration should use suitable standard buffers.

Data Analysis

For each site, I will calculate the mean conductivity and pH from the three replicates and report the spread, such as range or standard deviation. Individual values will remain visible in the laboratory record. A simple graph can compare mean conductivity by water-body type, but the final written article does not require a table.

If the differences are large and consistent among replicates, the results may support the hypothesis. If variation within one site is as large as variation among sites, the sampling design may be insufficient. A more advanced study could use more sites, several distances from the water, seasonal sampling, ion analysis, and statistical testing.

Laboratory and Field Safety

The main safety concern is not merely avoiding missing samples. Field collection near water creates risks of slippery banks, waves, tides, unstable sediment, sharp objects, contaminated water, heat, and traffic. Sampling should be performed with permission, appropriate footwear, gloves, awareness of weather and tides, and another person nearby where conditions are hazardous. No sample should be collected by entering deep or fast-moving water.

In the laboratory, samples should be treated as potentially contaminated. Gloves and eye protection should be used when necessary, hands should be washed, and work surfaces should be cleaned. Containers and instruments must be labeled clearly to prevent mixing. Electrical equipment should be kept dry, and calibration solutions should be handled according to their safety information. Soil should not be tasted to estimate salinity.

Expected Interpretation

If ocean soil produces the highest conductivity, the finding will be consistent with exposure to seawater or sea-derived salts. If river soil has the lowest reading, the result may reflect freshwater

conditions and leaching. Lake soil will require interpretation based on the lake's hydrology. The experiment can demonstrate local differences but cannot support a universal statement about every river, lake, and ocean.

Measured conductivity can be compared with published salinity classifications, but crop recommendations should consider the extraction method and plant tolerance. Soil management options may include drainage improvement, leaching with suitable water, organic matter, salt-tolerant crops, and prevention of saline irrigation. Applying more water without drainage can worsen the problem by raising groundwater (United States Salinity Laboratory Staff, 1954).

Conclusion

This experiment will compare soil salinity near a river, lake, and ocean through replicated sampling and electrical-conductivity measurement. The hypothesis predicts that coastal soil will be most saline, but geology, evaporation, drainage, flooding, and human activity may produce a different order. The study will also measure pH while recognizing that pH and salinity are separate properties.

The revised method corrects several problems in the original plan. Seawater salinity must be expressed with units; catfish do not meaningfully heat water; organisms do not create most ocean salt; and riverbank soil is not automatically fertile. Electrical conductivity of a saturated paste extract or a clearly reported fixed-ratio extract provides a defensible measurement.

The most important scientific practice is consistency. Samples must be collected at comparable depths and distances, processed through the same method, measured with calibrated instruments, and interpreted with local environmental information. The outcome will describe the selected sites and provide evidence for further study rather than prove a universal rule about every water body.

References

Corwin, D. L., & Yemoto, K. (2017). Salinity: Electrical conductivity and total dissolved solids. In *Methods of Soil Analysis*. Soil Science Society of America.

U.S. Department of Agriculture, Agricultural Research Service. (2017). *Laboratory measurement and monitoring of soil salinity*.

United States Salinity Laboratory Staff. (1954). *Diagnosis and improvement of saline and alkali soils*. U.S. Department of Agriculture.